

# Evaluation of the haematology and biochemistry of the silver catfish, *Chrysichthys nigrodigitatus* as biomarker of environmental pollution in a tropical lagoon

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#### ABSTRACT

The objective of the present study was to examine the haematological and biochemical parameters of the silver catfish, Chrysichthys nigrodigitatus, from the Makoko area of a polluted tropical lagoon such as Lagos Lagoon, to serve as a baseline data for the assessment of the health status of the fish and as reference point for future studies. The mean values of the haematological parameters analyzed were: blood haemoglobin, Hb (97.29±4.35g/L); pack cell volume, PCV (29.28±1.23%); red blood cell/total erythrocyte count, RBC (2.97±0.12T/L); white blood cell count, WBC (10.69±0.37g/L); mean cell volume, MCV (95.19±2.28fl); mean corpuscular haemoglobin, MCH (30.53±0.67pg), mean corpuscular haemoglobin concentration, MCHC (33.32±0.38g/dl), and the differential leucocytes. While the mean values of the biochemical parameters were: aspartate transaminase, AST (66.06±6.12 IU/L); alanine transaminase, ALT (12.98±1.10 IU/L), and alkaline phosphatase, ALP (88.01±8.60 IU/L). The findings from this study gave an indication of stress on the health status of the fishes in this lagoon and there is need for proper management strategies to be adopted in monitoring the conditions of the faunal communities in the lagoon. The lagoon indeed showed signs of environmental stress, which eventually pose devastating effect on the health status of the fishes. These have an indirect effect on the coastal communities. There is therefore the need for periodic monitoring and enforcement of environmental laws by respective organisations to ensure health safety of the people especially in the coastal communities.

Keywords: Haematological parameters; Chrysichthys nigrodigitatus; Makoko; Lagos Lagoon; baseline

# **INTRODUCTION**

Many of the developing countries of the world, such as Nigeria, aim at attaining the status of the developed nations in the nearest future. This goal is being achieved through massive industrialization and urbanization, which is leading to the continual pollution of the coastal waters. Akpata (1986) also emphasized that although aquatic pollution could be attributed to inadequate consideration given to environmental impact analysis of various projects; it is majorly due to increasing urbanization and industrialization of the country's coastal cities. According to Babalola and Agbebi (2013), water pollution is a major problem in the global context, but Lagoon pollution has been increasingly significant over the recent years and this has been found to contribute significantly to environmental problems in many developing countries where, through man's exploitation of the water resources, the normal dynamic balance in the aquatic ecosystem is continuously disturbed, and often results in each dramatic response as depletion of fauna and flora, fish kill, and change in physico-chemical character. The Makoko area of Lagos Lagoon is being used as the cheapest and convenient refuse disposal system. The indiscriminate discharge of wastes into the coastal waters has resulted in excessive loading of these aquatic systems, beyond their capacity of self-purification.

*C. nigrodigitatus* was selected for this particular study because of its relative availability and abundance of the fish species throughout the year in the Lagos Lagoon, and widely consumed by the coastal communities and sold in several markets in the area (Fagade and Olaniyan, 1973; 1974). It is a mainstay of the artisanal fisheries in coastal communities in the area. It has great aquaculture

potentials (Oribhabor and Ezenwa, 2005), and thus several studies have been carried out on the species (Ekanem, 2000; Offem *et al.*, 2008; Nwafili *et al.*, 2012).

The natural environment of fish is water. They live in direct contact with water. Changes in the physico-chemistry of water do induce physiological stress to the fish. In order to survive in its environment, the fish would have to either overcome the stress or adapt to it. The physiological stress response, although initiated as an adaptive response to destabilizing factors, could have damaging effects if prolonged. Jobling and Reinsnes (1986) established that continuous stress affects the behaviour and normal development of fish. Gerking 1980) observed that these stressors caused growth reduction, while Schreck and Bradford (1990) stated that the stressors were responsible for suppression of reproduction and an increase in susceptibility to infections, through immune depression, which caused mortality. Therefore, there has been a greater understanding of the need to establish reference haematological and biochemical values in fish in order to assess health status and the subsequent diagnosis of disease. A similar study was carried out for fish species reared in ponds by other researchers, for example by Etim *et al.* (1999), Sowunmi (2003). Erondu *et al.* (1993); Adedeji *et al.* (2000), and Gabriel *et al.* (2004).

Haematological parameters such as RBC (red blood cells), Hb (haemoglobin), Haematocrit/packed cell volume (HCT/PCV), MCV (mean corpuscular volume), MCH (mean corpuscular haemoglobin), and MCHC (mean corpuscular haemoglobin concentration); and biochemical parameters such as AST (aspartate aminotransferase), ALT (alanine aminotransferase) and ALP (alkaline phosphatase), are the most common criteria used in the toxicity studies on fish, and the present study was to examine these parameters in the catfish, *C. nigrodigitatus* from a contaminated tropical water body as Lagos Lagoon. Hence, the objective of the present study was to examine the haematology and biochemical parameters of the silver catfish, *C. nigrodigitatus* from a tropical polluted Lagos Lagoon, as a biomarker of the health status of the species.

### MATERIALS AND METHODS

#### **Study Site**

Makoko is a slum neighbourhood community located on the eastern part of Lagos, Nigeria (Figure 1). According to Ayeni (2014), Makoko had an estimated population of about 85,000 people in 2009. Majority of these people engage in fishing as means of livelihood. Over a thousand fishing boats (paddled and motorized) could be located at a time.

#### **Fish Species**

Silver catfish, *Chrysichthys nigrodigitatus* (Lacépède: 1803),of the family Claroteidae plays a pivotal role in the ecology and fisheries of Nigeria in particular, and West Africa at large (Ayotunde and Ada, 2013).*C. nigrodigitatus* is an important commercial fish because of its high protein content and hardy flesh, thus forming a very important component in the diet of many Nigerians. According to Ezenwa *et al.* (1990), *C. nigrodigitatus* accounts for the second highest production of fish that can be obtained from the wild in the four geomorphological regions of the Nigerian coastline, and is highly recommended as a culturable species. In the Makoko area, *C. nigrodigitatus* is the second most landed fish species, after the various tilapia species. The fishes were landed through the help of the local fishermen using cast nets of various mesh sizes. The weight ranged between 1000 - 1420 g, while the length ranged from 31 - 51 cm.



Figure 1. Map of Lagos Lagoon showing the Makoko Slum Area

# **Blood Sampling**

A bi-monthly collection of blood from 18 live fish specimen was carried out between May and July, 2013. The bloods were collected between 08:00 and 10:00 hour on each of the sampling day, stored in ice chest and transported to the laboratory for analyses. The blood collection was done using 2ml sterile plastic disposable syringes fitted with  $0.8 \times 38$ -mm hypodermic needles. The blood samples were expressed into ethylene diamine tetra acetic acid (ETA) and lithium heparinized bottles for the haematology and serum enzyme biochemical analyses respectively.

# Haematological and Biochemical Procedures

Haemoglobin (Hb) concentration was determined with the cyanomethaemoglobin method. Packed Cell Volume (PCV) by micro haematocrit method, Red blood cell (RBC) and total white blood cell (WBC) were done using the Neubauer haemocytometer. Differential counts (neutrophils, monocytes and lymphocytes) were done on blood film stained with May Grumwald-Giensa stain. The mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC) were calculated from the data using standard formulae.

Serum was separated from the cellular blood components by centrifugation for 5 min at 14,000 rev/min. Blood alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (AP) were determined using a portable automated chemical analyzer following the procedure and using the reagents recommended and as described by the manufacturer (RANDOX Laboratories Ltd, UK; AST/ALT (Cat. No. Sc 2643) and ALP multi-sera level 2 (Cat. No. 1530) and level 3 (Cat. No. 1532)), assay kits. All blood analyses were carried out within 48 hours of collection.

### Statistical Analyses of Data

Haematological parameters and biochemical indices were analysed using one-way analysis of variance (ANOVA) at 5% level of significance. While post-hoc comparison of significance of variance result gotten from ANOVA was done using Duncan Multiple Range Test (DMRT).

### RESULTS

The analysis of variance (ANOVA) of the haematological parameters showed that only Hb, PCV and WBC are significant (P<0.05) while Post-Hoc analysis using DMRT (Duncan Multiple Range Test) showed that Hb, PCV, WBC and MCV are significant (P<0.05). But analysis of variance of the biochemical parameters showed that there was significant difference (P<0.05) in all of them, while Post-Hoc analysis using DMRT showed that there was significant difference in AST in weeks 2, 3 and 6; ALT in weeks 2, 3, 4 and 5; ALP in weeks 2, 3, 4 and 5. The results of the haematological and biochemical analyses are shown in Tables 1 and Table 2, respectively.

Table 1. Mean and S.E for Haematology of Chrysichthys nigrodigitatus

Parameters	Week-1	Week-2	Week-3	Week-4	Week-5	Week-6	Grand Mean
Hb(g/L)	$101.7 \pm 0.98^{b}$	99.23±5.15 <sup>b</sup>	$63.37 \pm 5.56^{a}$	101.83±8.15 <sup>b</sup>	104.83±6.64 <sup>b</sup>	$112.8 \pm 7.2^{b}$	97.29±4.35
PCV (%)	$29.67 \pm 0.88^{b}$	$30\pm0.58^{b}$	19.67±2.19b	$31.67 \pm 2.40^{b}$	31.33±1.76 <sup>b</sup>	33.33±1.45 <sup>b</sup>	29.28±1.23
RBC(T/L)	3.13±0.07 <sup>a</sup>	$3.23 {\pm} 0.15^{a}$	$2.37 \pm 0.32^{a}$	3.27±0.44ª	3.13±0.24 <sup>a</sup>	$2.67 \pm 0.29^{a}$	$2.97 \pm 0.12$
WB(g/L)	12.79±0.66 <sup>b</sup>	$11.51 \pm 0.44^{ab}$	10.11±1.06ª	9.31±0.50ª	$10.59 \pm 0.62^{ab}$	9.83±0.66 ª	$10.69 \pm 0.37$
MCV(fl)	$92.94 \pm 0.46^{ab}$	90.26±0.87ª	92.37±7.40 <sup>ab</sup>	91.20±3.10ª	$95.72 \pm 5.04^{ab}$	$108.67 \pm 7.5^{\mathrm{b}}$	95.19±2.28
MCH(pg)	$32.13 \pm 0.13^{b}$	$30.54 \pm 0.60^{\text{b}}$	31.54±3.37b	29.24±0.56b	$31.48 \pm 0.74^{\mathrm{b}}$	$28.27 \pm 2.21^{b}$	30.53±0.67
MCHC(g/dl)	33.60±1.84ª	33.25±1.80ª	33.10±0.20ª	33.40±0.32ª	33.62±0.47ª	$32.97 \pm 0.19^{a}$	33.32±0.38
NEUT(%)	$13.67 \pm 2.19^{a}$	18.67±4.67ª	$19.00 \pm 5.29^{a}$	$20.67 \pm 1.45^{a}$	17.33±1.20ª	$21.67 \pm 1.20^{a}$	18.50±1.25
LYMP(%)	86.33±3.18ª	74.33±7.22ª	84.67±2.19ª	$81.67 \pm 2.60^{a}$	82.67±1.45ª	$78.33 \pm 2.84^{a}$	81.33±1.61
MONO(%)	$0.67 \pm 0.67$ a	$1.00 \pm 1.00^{a}$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.33 \pm 0.33^{a}$	$0.00 \pm 0.00$	0.33±0.20
EOS(%)	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$
BAS(%)	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$	$0.00 \pm 0.00$

Mean frequencies (mean  $\pm$  S.E, Standard Error) with different superscript letters in a row are significantly different in the DMRT (p<0.05). NEUT (neutrophile granulocytes); LYMP (lymphocytes); MONO (monocytes); EOS (eosinophile granulocytes); BAS (basophile granulocytes)

Table 2. Biochemical Parameter of Chrysichthys nigrodigitatus											
Parameters	Week-1	Week-2	Week-3	Week-4	Week-5	Week-6	Grand Mean				
AST(IU/L)	$36.07 \pm 5.85^{a}$	$91.09 \pm 0.80^{b}$	81.86±10.87b	53.33±3.48ª	39.43±3.79ª	94.59±0.63b	66.06±6.12				
	0.0414.40										
ALT(IU/L)	$8.96 \pm 1.60^{a}$	14.51±1.21 <sup>b</sup>	21.06±0.44 <sup>c</sup>	12.56±1.20 <sup>b</sup>	$13.07 \pm 0.05^{b}$	$7.73 \pm 0.14^{a}$	$12.98 \pm 1.10$				
ALP(IU/L)	$33.12 \pm 4.22^{a}$	125.12±6.63 <sup>d</sup>	119.6±3.68 <sup>cd</sup>	103.04±13.55 <sup>bc</sup>	95.68±2.43 <sup>b</sup>	$51.52 \pm 1.84^{a}$	88.01±8.60				

Mean frequencies (mean  $\pm$  S.E, Standard Error) with different superscript letters in a row are significantly different in the DMRT (p<0.05)

### DISCUSSION

This present study sought to provide haematological indices and aspects of biochemical indices for C. nigrodigitatus in Makoko area of a tropical and polluted Lagos Lagoon, a brackish water environment. Okafor and Chukwu, (2010) also confirmed the need to establish normal haematological characteristics of a particular species of fish which would serve as reference for further comparative studies. This is very necessary, as emphasized by Moiseenko (1998), and Adhikari et al. (2004) that haematological assessment is a pathophysiological reflector of the whole body and, therefore, blood parameters are important in diagnosing the structural and functional status of fish exposed to contaminants. From the report by Adedeji and Adegbile (2011), on the haematology of C. nigrodigitatus from a freshwater Asejire Dam in Nigeria, all the blood parameters such as PCV, Hb, RBC, WBC and Lymphocytes were lower than those in the present study. According to Hassan et al., (2016), increase in salinity gradient was noted to have an increasing effect on the value of the blood parameters in the common carp, Cyprinus carpio. These values are in line with those of Okafor and Chukwu (2010) who recorded mean Haematocrit values of 27.7%, 28.1%, 28.8% and 29.2% for the fingerlings, juveniles, intermediates and large sizes, respectively, for the African Lungfish (Protopterus annectens) from Anambra River. The values recorded in this study are also within the minimum, intermediate and maximum values of 20%, 35% and 50% respectively, in various reports (Clarks et al., 1979; Etim et al., 1999). According to Adedeji and Adegbile (2011), anaemic condition is determined by haematocrit or PCV (Blaxhall and Daisley, 1973); increased Hb concentration depict higher activity; increased RBC also depicted more effective O2 and CO2 to lung tissue; while WBC and Lymphocytes increase is an indication of ability to fight infections from the environment. However, it is surprising that a fish in a polluted environment as the Lagos Lagoon would have increased blood parameters as should be in a healthy fish. This could possibly be as an internal homeostatic response in the C. nigrodigitatus to counteract the stresses posed both by a saline gradient and the pollutants in the Lagoon.

Toxic effects of different pollutants are frequently determined by the use of Plasma enzymes, AST, ALT, and LDH (Li *et al.*, 2010). The biochemical parameter in the present study, AST, was higher than the mean value of  $48.26\pm77.54$  reported for *C. nigrodigitatus* by Adedeji and Adegbile (2011). However, ALT value in this study was lesser than that reported (21.79±13.49) by Ayoola *et al.* (2014). Increase in AST occurred due to disruption in mitochondria brought about as a result of heavy hepatitis; in the same way, elevated level of ALT in the blood is indicative of damage to the integrity of hepatocyte membranes (Ayoola *et al.*, 2014).

The comparable levels of haematological parameters in this study could also be attributed to the various stressors present in the Lagoon. Various authors, such as Gabriel *et al.* (2007), have established that pollutants, such as herbicides, pesticide and industrial effluents, alter the haematological indices of fish. Makoko Area of Lagos Lagoon is organically polluted, especially with sewage from the floating shanties, wood wastes from logging and sawmills activities and Refuse discharges from the nearby Better-life fish market. There was disparity in the colour of fish species; while some of the *C. nigrodigitatus* were silver-white; others were silver-grey, which could also be an evidence of pollution. Sarkar and Bhavna (2011) also reported that pollution is capable of changing the colour/physical appearance of a fish.

# CONCLUSIONS

The lagoon indeed showed signs of environmental stress, which eventually pose devastating effect on the health status of the fishes. These have an indirect effect on the coastal communities. There is therefore the need for periodic monitoring and enforcement of environmental laws by respective organizations to ensure health safety of the people especially in the coastal communities.

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#### REFERENCES

- Adedeji, O.B., A.F. Adegbile. 2011. Comparative haematological parameters of the bagrid catfish (*C. nigrodigitatus*) and the African catfish (*Clarias gariepinus*) from Asejire Dam in Southwestern Nigeria. Journal of Applied Sciences Research, 7(7): 1042-1046.
- Adedeji, O.B., O. Taiwo, S.A. Agbede. 2000. Comparative haematology of five Nigerian freshwater fish species. Nigerian Veterinary Journal, 21: 75–84.
- Adhikari, S., B. Sarkar, A. Chatterjee, C.T. Mahapatra, S. Ayyappan. 2004. Effects of cypermethrin and carboforan on certain haematological parameters and prediction of their recovery in a fresh water teleost, *Labeo robita* (Hamilton). Ecotoxicology and Environmental Safety, 58:220-226.
- Akpata, T.V.I. 1986. Pollution: Flora of some wetlands in Nigeria. In: Nigerian wetlands. Akpata, T.V.I. and Okali, D.V.V. (Eds.). The Nigerian Man and the Biosphere National Committee 1990. Emmi Press. Somonda, Ibadan. 198pp.
- Ayeni, A.O. 2014. Domestic water source, sanitation and high risk of bacteriological diseases in the urban slum: Case of cholera in Makoko, Lagos, Nigeria. Journal of Environment Pollution and Human Health, 2(1): 12-15.
- Ayoola, S.O., S.C. Akagha, K.O. Ezeanyika. 2014. Plasma and enzymatic indices of *C. nigrodigitatus* at the Bariga landing site, Lagos Lagoon. PAT, 10(1): 151-163.
- Ayotunde, E. O., F.B., Ada. 2013. Silver catfish, *Chrysichthys nigrodigitatus* (Lacépède: 1803), an endangered fish species in Cross River, Cross River state, Nigeria. International Journal of Agricultural Science Research, 2(3): 83-89.
- Babalola, O.A., F.O. Agbebi. 2013. Physico-chemical characteristics and water quality assessment from Kuramo Lagoon, Lagos, Nigeria. International Journal of Advanced Biological Research, 3(1): 98-102.
- Blaxhall, P.C., K.W. Daisley. 1973. Routine haematological methods for use with fish blood. Journal of Fish Biology. 5: 771-781.
- Clarks, S., D.H. Whitemore, Jr. R.F. McMahon. 1979. Consideration of blood parameters of largemouth bass, Micropterus salmoides. Journal of Fish Biology, 14:14 –158.
- Ekanem, S.B. 2000. Some reproductive aspects of *Chrysichthys nigrodigitatus* (Lacepede) from Cross River, Nigeria. Naga, ICLARM Quarterly, 23(2):2 4-28.
- Erondu, E.S., C. Nnubia, F.O. Nwadukwe. 1993. Haematological studies on four catfish species raised in freshwater ponds in Nigerian Journal of Applied Ichthyology, 9:250–256.
- Etim, L., S.B. Ekanem, A. Utin. 1999. Haematological profile of two species of catfish, *Chrysichthys nigrodigitatus* (Lacepede) and *Chrysichthys furcatus* (Gunter) from the Great Kwa River, Nigeria. Global Journal of Pure and Applied Science, 5: 1-4.

- Ezenwa, B.I.O., W.O. Alegbeleye, P.E. Anyanwu, E.O. Uzukwu. 1990. Culturable fish seeds in Nigerian Coastal Waters. A research survey (Second Phase: 1986-1989). Nigerian Institute for Oceanography and Marine Research (N.I.O.M.R.) Tech paper 37. 66pp.
- Fagade, S.O., C.I.O. Olaniyan. 1973. The food and feeding interrelationships of fishes in Lagos Lagoon. Journal of Fish Biology ,5:205-225.
- Gabriel, U.U., E.U. Amakiri, G.N.O. Ezeri. 2007. Haematology and gill pathology of *Clarias gariepinus* exposed to refined petroleum kerosene under laboratory conditions. Journal of Animal and Veterinary Advances, 6(3): 461-465.
- Gabriel, U.U., G.N.O. Ezeri, O.O. Opabunmi. 2004. Influence of sex, health status and acclimation on the haematology of Clarias *gariepinus* (Burchell, 1822). African Journal of Biotechnology, 3(9):463–467.
- Gerking, S.D. 1980. Fish reproduction and stress. In: Environmental physiology of fishes. Plenum Press, New York. 406pp.
- Hasan, A. Al Hilali, S. Mohammed, Al-Khshali. 2016. Effect of water salinity on some blood parameters of common Carp (*Cyprinus carpio*). International Journal of Applied Agricultural Sciences, 2(1): 17-20.
- Jobling, M., T.C. Reinsnes. 1986. Physiological and social constraints on growth of Arctic charr. *Salvelinu salpinus* (L): an investigation of factors leading to stunting. Journal of Fish Biology, 28:379-384.
- Moiseenko, T.I. 1998. Haematological indices of fish in the evaluation of their toxicosis with reference to *Coregonus lavaretus*. Journal of Ichthyology, 38:315-324.
- Nwafili, S.A., O.O. Soyinka, T.X. Gao. 2012. Levels and patterns of genetic diversity in wild *Chrysichthys* nigrodigitatus in the Lagos Lagoon complex. African Journal of Biotechnology, 11(91): 15748-15754.
- Offem, B.O., Y. Akegbejo-Samsons, T.I. Omoniyi. 2008. Diet, size and reproductive biology of the silver catfish, *Chrysichthys nigrodigitatus* (Siluriformes: Bagridae) in the Cross River, Nigeria. International Journal of. Tropical Biology, 56(4):1785-1799.
- Okafor, A.I., L.O. Chukwu. 2010. Haematological profile of the African lungfish, *Protopterus annectens* (Owen) of Anambra River, Nigeria. Journal of American Science, 6(2):123-130.
- Oribhabor, B., B. Ezenwa. 2005. Inventory of fisheries and fishes of the Lagos Lagoon, Lagos, Nigeria. Tropical Freshwater Biology, 14:19-36.
- Schreck, C.B., C.S., Bradford. 1990. Internal corticosteroid production: potential regulation by immune system in the salmonids. Progress in Clinical and Biological Research, 342:480-486.
- Sowunmi, A.A. 2003. Haematology of the African catfish, *Clarias gariepinus* (Burchell, 1822) from Eleiyele reservoir, Ibadan, Southwest Nigeria. The Zoologist, 2(1): 40–44.

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